



## Raji

CSI179Hu11

*Instruction manual*

FOR RESEARCH USE ONLY

NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

2nd Edition (Revised in Apr, 2025)

### [ DESCRIPTION ]

The Raji line of lymphoblast-like cells was established by R.J.V. Pulvertaft in 1963 from a Burkitt's lymphoma of the left maxilla of an 11-year-old Black male. This cell line can be used in immunology research.

**Synonyms:** RAJI; P1-Raji

**Organism:** Homo sapiens, human

**Tissue Source:** Burkitt's lymphoma

**Disease:** Burkitt's lymphoma

**Age:** 11 years

**Gender:** Male

**Cell Type:** B lymphocyte

**Morphology:** lymphoblast

**Growth properties:** Suspension

### [ PROPERTIES ]

**Cell activity:** >85% (Viability by Trypan Blue Exclusion).

**Formulation:** Frozen 1 mL or T25 flask.

**Biosafety:** Negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi.

**Applications:** For research use only. It is not approved for human or animal use, or for application in clinical diagnostic procedures.

**Size:**  $>5 \times 10^5$  cell/vial

### [ STORAGE ]

Upon receiving, check all containers for leakage or breakage. directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

**Form & Buffer:** Supplied as solution form in frozen stock solution, containing 50% base medium +40%FBS+10%DMSO.

**Storage conditions:** liquid nitrogen

### [ USAGE ]

**Culture conditions:**

Complete growth medium: RPMI-1640+10%FBS+1%Penicillin-Streptomycin Solution



Temperature: 37°C

Condition: 95% air, 5% carbon dioxide

**Cell recovery:**

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. The thawing time is about 2 minutes.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 75% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0mL complete culture medium. and spin at approximately 1000 rpm for 5 minutes.
4. Resuspend cell pellet with the recommended complete medium . and dispense into a T25 culture flask.
5. Incubate the culture at 37°C, 5% CO<sub>2</sub> in a suitable incubator.

**Cell passage:**

The cells are suspended cells, and maintain cultures at a cell concentraion between  $4 \times 10^5$  and  $3 \times 10^6$  viable cells/mL. Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at  $4 \times 10^5$  viable cells/mL, and centrifugal speed reference 1200 rpm centrifuge 3 min. According to culture experience, it is recommended to use the "half-change solution method" for passage, that is, directly add an equal amount of fresh culture solution to the cell culture bottle, then the cells are blown evenly and transferred to two new T25 culture bottles for further culture. Depending on cell density, it is recommended to add fresh medium every 2-3 days.

## [ Shipping ]

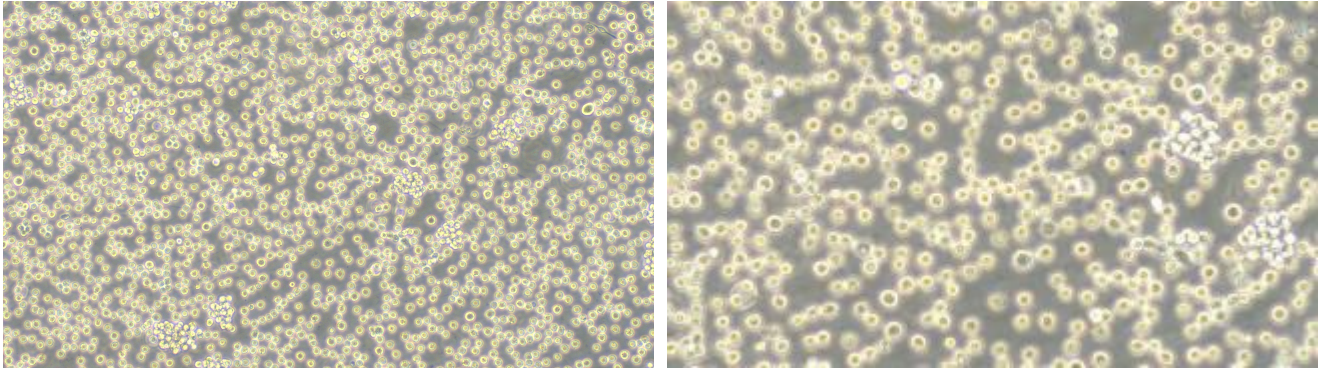
Dry ice.

## [ IMPORTANTNOTE ]

1. This product is intended for laboratory research use only. It is not intended for any animal or human therapeutic use, any human or animal consumption, or any diagnostic use.
2. To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.
3. After cell recovery, please take regular microscopic examination and photos to record the growth status of cells.
4. Read the instructions carefully, and keep and operate in strict accordance with the instructions. If you observe abnormalities or have questions about cell culture operations, please contact us in time.



**[ Figure ]**



Morphology of Raji (Optical microscope, 100x, 200x)